

Fresh Crushed Garlic Exhibits Superior Allicin and Pyruvic Acid Stability, While Fresh Sliced Garlic Leads in Phenolic and Antioxidant Content

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Running Title: Peak Allicin Content, Antioxidant Activity, and Pyruvic Acid Levels....

Keywords: Allicin, Antioxidants, Nutrition, Drying, Storage.

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Abstract

This study investigates the stability of allicin, phenols, and antioxidants in different forms of garlic (fresh whole peeled, fresh sliced, fresh crushed, and dried slices) under various storage conditions (0 to 2 days at 4 °C and 20 °C). Fresh garlic exhibited allicin levels ranging from 0.6 to 32.14 mg/g, while dried garlic showed significantly lower levels (3.77 to 6.68 mg/g). Maximum allicin stability in fresh garlic was observed after 10 minutes at 20 °C, with pyruvic acid peaking after 10 minutes at 4 °C. Freshly sliced and crushed garlic retained the highest phenol content and antioxidant activity immediately after preparation, whereas dried garlic had reduced levels due to thermal processing. Consumer testing revealed that hummus with fresh crushed garlic at 10 minutes and 20 °C was preferred, while dried garlic at 20 minutes and 4 °C had the strongest flavor and aroma. In conclusion, fresh crushed garlic is optimal for allicin and pyruvic acid content, while fresh sliced garlic excels in phenol and antioxidant levels; drying notably diminishes these beneficial compounds, affecting flavor and consumer preference.

INTRODUCTION

Garlic, *Allium sativum*, is one of the ancient crops, and it has been named the "aroma" vegetable since it has been used as a flavoring in cuisine and as a medication due to its numerous health and wellness advantages in many civilizations (Botas *et al.*, 2019; Netzel, 2020). Garlic has a high concentration of functionally active compounds and a diverse chemical composition (Marchese *et al.*, 2016). Multiple research studies have demonstrated that the health benefits attributed to fresh garlic extract are believed to be associated with the presence of a total of 33 sulfur compounds (Amagase, 2006; Bhandari, 2012; Kovarovič *et al.*, 2019). Those benefits include antioxidant properties, anti-inflammatory activity, antimicrobial activity, immunological activation,

cardiovascular defense, antitumor, anti-carcinogenic, gastric safety, anti-diabetic function, anti-obesity activity, the nervous and endocrine systems (Shang *et al.*, 2019; Gam *et al.*, 2021).

Garlic has a weak, undetectable odor until it is peeled. When peeled, sliced, or crushed, it forms a compound called allicin, which releases a strong odor containing sulfur glycosides (Yusuf *et al.*, 2018). Allicin, or diallyl thiosulfates, is the most active biological ingredient in garlic (Farías-Campomanes *et al.*, 2014), accounting for over 70% of the total thiosulfates found in cloves (Rahman, 2007), it's highly unstable and can breakdown even at ambient temperature (Ilić *et al.*, 2015; Zhu and Zeng, 2020), in a matter of days or hours to far more stable sulfur-containing compounds (Rybak *et al.*, 2004). Furthermore, allicin is not present in garlic; it is synthesized from alliin (S-Allyl-L cysteine-sulfoxides), an odorless derivative of cysteine, by the enzyme allinase (Abe *et al.*, 2020). Upon processing or consumption, these compounds contribute to the distinct pungent aroma of garlic (Kovarovič *et al.*, 2019).

Khar *et al.*, (2011) studied 93 garlic ecotypes for allicin content and observed variability in allicin content in each one. Prati *et al.*, (2014) studied allicin content in 5 cultivars for three forms of processed garlic (unsalted garlic paste, chopped fried garlic, and fried sliced garlic) for a long period of storage reaching 180 days, with the most loss of allicin recorded in fried garlic, followed by paste. Another researcher reported that cooking methods at home (boiling, microwaving, pressure cooking, griddling, frying, and baking) have been demonstrated to affect the important components and antioxidant activity of garlic (Martins *et al.*, 2016). The stability of allicin, which is typically thought of as unstable in nature, has been the subject of several research. The stability of allicin varies depending on environmental parameters including temperature and the solvent used to dissolve it (Chan *et al.*, 2012). This unstable chemical has poor solubility in water (Marchese *et al.*, 2016). The most prevalent bioactive components of garlic are naturally sensitive

to temperature degradation, which reduces their effectiveness (Martins *et al.*, 2016). The high volatility and instability of allicin limit the use of garlic as a pharmacologically active drug, which significantly restricts its commercial usage. Also, Allicin's instability is one issue that inhibits its chemical uses (Ilić *et al.*, 2011).

Given the significant volatility of organosulfur compounds in garlic, such as allicin, proper storage conditions are critical for retaining the excellent quality of garlic bulbs and by-products. The period of storage is critical for garlic's bioactive qualities (Martins *et al.*, 2016). Also, storage temperature is particularly significant since it can influence the chemical structure and, as a result, the ultimate bioactivity intensity of garlic and allicin content over time (Veríssimo *et al.*, 2010). Most of the previous work available in the literature relates to the evaluation of allicin at high temperatures, long storage interval periods (up to 45 days or 180 days), and the comparison of different ecotypes or cultivars of garlic. From another point of view, for processed foods, the approach has a substantial impact on the retention of bioactive chemicals from garlic, especially those connected to antioxidant properties like the one achieved by allicin from garlic (Prati *et al.*, 2014). The previous work available in the literature relates to the evaluation of allicin stability in different solvent extractions, showing allicin stability in polar and nonpolar solvents after extraction (Zalepugin *et al.*, (2015). In our study, we evaluated allicin stability before the extraction process in different forms (freshly sliced, freshly crushed, and dried slices) during short storage intervals, which were considered practical for daily use. Based on the above, the aims of this study are to evaluate the stability of allicin in fresh sliced garlic, freshly crushed garlic, and dried garlic slices under different storage conditions of time and temperature (from zero time to two days at 4 °C and 20 °C).

MATERIALS AND METHODS

97 **Materials**

98 Fresh white garlic (*Allium sativum* L.) bulbs and hummus (a dip made from pureed chickpeas
99 sesame seed paste and lemon) were purchased from the local market in Irbid, Jordan. Allicin
100 standard (purity 98%) was purchased from (TargetMol, EU). All chemical reagents utilized in this
101 study were purchased from Sigma-Aldrich (St. Louis, MO, USA) or Fisher Scientific (Pittsburgh,
102 PA, USA).

103 **Sample Preparation**

104 This study analyzed three different forms of garlic: freshly sliced, freshly crushed, and dried garlic
105 slices. Fresh whole garlic was used as a control. Thirteen samples were prepared, each weighing
106 10 g, and stored under different conditions. Fresh garlic was cut into uniform slices (3.5 mm) using
107 a kitchen knife (Fresh sliced), crushed into a paste using a garlic press (Fresh crushed), and dried
108 slices were dried in the oven at 60 °C for 24 hours (Dried garlic). The garlic samples were prepared,
109 labeled, and stored at different temperatures for different times as described in Table 1.

110 **Garlic Extraction**

111 Garlic was extracted according to the method described by Bat-Chen *et al.*, (2010) with some
112 appropriate modifications. The residue was reconstituted with 5 ml of the mobile phase (50 %
113 Water: 50 % Methanol) and homogenized by placing it in a closed beaker, then sonicated before
114 filtration.

115 **Identification of Allicin in Garlic Extract by HPLC**

116 Allicin stock solutions were prepared with 50 mg of allicin (Commercially available) dissolved in
117 5 ml of 60:40 (mixtures of water and methanol) as described by Rybak *et al.*, (2004). The

118 identification of the allicin peak was at 220 nm wavelength. Two vials were injected in the HPLC
119 to determine the allicin content in the samples. The first vial was garlic extract without the
120 standard, and the second was garlic extract with the standard. Then, the intensity of the peak was
121 compared to the standard.

122 **Determination of Allicin Content in Garlic Extract by HPLC**

123 The identification and quantification of allicin were performed in a high-performance liquid
124 chromatography (HPLC) system (SHIMADZU, Japan). The determination of allicin in the garlic
125 sample was performed according to the procedure described by de Diego et al., (2007) with some
126 modifications in column dimension (100 mm x 3 mm).

127 **Determination of Pyruvic Acid Content in Garlic**

128 The pyruvic acid content in garlic extract was measured according to a procedure described by
129 Anthon and Barrett, (2003).

130 **Determination of Total Phenol Content**

131 The total phenol content of the garlic extract was determined using the Folin–Ciocalteu method,
132 as previously described by Alu'datt *et al.*, (2010).

133 **Determination of Antioxidant Activity**

134 Anti-oxidant activity of the garlic extract was determined using a method described by (Brand-
135 Williams *et al.*, 1995).

136 **Sensory Evaluation**

137 The study was reviewed and approved by the scientific research committee at the Department of
138 Nutrition and Food Technology, Jordan University of Science and Technology to perform the

139 sensory evaluation. The samples are hummus with different treatments of garlic prepared
140 according to the method conducted by Olaimat *et al.*, (2017). A sensory evaluation of hummus
141 samples was conducted using a hedonic scale of acceptance (Yilmaz and Aydeniz, 2012), and Just
142 About Right (JAR) scales to assess whether the intensity of an attribute is appropriate (Lawless
143 and Heymann, 2010). A total of 40 participants from Jordan University of Science and
144 Technology, aged between 18 and 30 years participated in this study. Different samples of hummus
145 were prepared and garlic from different treatments was added to the hummus samples. Participants
146 were directed to taste samples and evaluate them at individual tables, each participant was provided
147 with thirteen plates. To eliminate carry-over factors, participants were also provided with green
148 apples and water for mouth cleaning between samples. The participants were asked to record their
149 acceptability scores (Hedonic scale) of samples for overall appearance, overall color, overall
150 aroma, overall pungency, and overall flavor (9-point scale with 9 = extremely like and 1 =
151 extremely dislike). Acceptability intensity scores (Just About Right (JAR) scales) of samples for
152 flavor, aroma, and pungency (5-point scale with 5 = normal and 1 = too much).

153 **Statistical Analysis**

154 Results were analyzed as the mean and standard deviation using the general linear model (GLM)
155 method with JMP statistical package (JMP Institute Inc., Cary, NC, USA). Means were isolated
156 using the least significant difference (LSD) test procedure. Statistical difference was considered
157 significant if $p\text{-value} \leq 0.05$.

158 **RESULTS AND DISCUSSION**

159 **Identification of Allicin in Garlic Extract by HPLC**

The main peaks in the chromatograms of garlic extract (GE) without standard and GE with standard are shown in **fig 1** and **fig 2**. The peak of allicin was identified by comparing peaks for GE without a standard in chromatograms with peaks for GE with a standard in chromatograms. So, the peak that increased with the increase in the amount of allicin (standard) is the peak that expresses allicin which was observed at retention time between 6 and 7 min (approximately 6.5 min).

Determination of Allicin Content in Garlic Extract by HPLC

Tables 2 and 3 demonstrate the mean of allicin content in fresh (sliced and crushed) and dried garlic slices with the effect of different storage conditions (time and temperature). The allicin contents were significantly varied ($p < 0.05$) among treatments and forms. The highest allicin content between all treatments is (32.14 mg/g) and was found in fresh crushed garlic in Trt 3 (20 °C, 10 min), whereas, the lowest allicin content (0.6 mg/g) was also found in fresh crushed garlic in Trt 13 (20 °C, 2 days). According to several studies, there is significant variability in the garlic content of organic sulfur profiles between previous studies and our study due to two factors. The first cause was the use of different extraction solvents, which affects the degradation of allicin into other compounds such as dichloromethane. The second is an analysis of the many garlic genotypes utilized in the tests (Martins *et al.*, 2016; Liu *et al.*, 2020).

Table 2 shows that the amount of allicin in fresh (sliced and crushed) garlic in Trt 1 (20 °C, time 0) is low because the enzymatic reaction that converts alliin to allicin takes about 10 minutes from the moment it is cut. The concentration of allicin in sliced and crushed fresh garlic increased after that in the following treatments, Trt 2 (4 °C, 10 min) and Trt 3 (20 °C, 10 min), and then the concentrations of allicin in the treatments began to decrease. These results match findings by Nguyen *et al.*, (2021) who found that the allicin content was around 8.3 mg/g at the first

measurement (0.5 min), but after 5 minutes, the allicin concentration rose significantly. When we compared the treatments of fresh sliced garlic and fresh crushed garlic, we found that the amount of allicin in the first form is less, but it is more stable. This corresponds to Thuwapanichayanan *et al.*, (2014), who confirmed that fresh garlic slices did not create as much allicin as crushed or diced garlic because allinase did not come into contact with alliin as frequently. The result, the content of allicin decreased with increasing time and temperature of storage, and this shows a relation between the decrease in stability of allicin with increased temperature and time of storage, and the opposite is true. these results agree with Fujisawa *et al.*, (2008), who found that the different conditions of temperature and time affect the stability of allicin, which is considered unstable. Table 3 shows that when we compare treatments of fresh sliced garlic with dried garlic slices, the allicin content of the first form is much higher. So, the shift in enzyme activity during the drying process may be responsible for the high reduction of allicin in dried garlic slices. Also, high temperatures may accelerate the degradation of allicin, which has been reported to be a heat-sensitive compound whose subsequent evaporation during drying was responsible for the loss (Zhou *et al.*, 2016). Our results in dried treatments were within the range reported by Rahman *et al.*, (2009), who showed allicin content in dried garlic is 4.932 mg/g. While drying garlic powder causes significant changes to the allinase, it still can convert alliin to allicin. The decrease of allicin in all treatments may be due to the natural variability of the degradation phenomenon and may also be attributed to the unstable allicin hydrolyzed and the decomposition to mercaptans, disulfides, trisulfides, and thiophenes (Henríquez *et al.*, 2014).

Pyruvic Acid Content

The mean of pyruvic acid content in all treatments is given in tables 4 and 5. The remaining flavor aptitude is represented by enzymatic pyruvic acid, as reported by Ammarellou *et al.*, (2022). The

results in tables 4 and 5 show that the content of pyruvic acid in the treatments differs according to the form of garlic (fresh sliced, fresh crushed, or dried slices). The pyruvic acid content varied substantially across all the treatments, with the highest value being 30.77 $\mu\text{mol/g}$ (wet basis) in fresh crushed garlic in Trt 2 (4 °C, 10 min storage). Based on the results of our study, the highest values of pyruvic acid were found in fresh crushed garlic, followed by fresh sliced, and dried slices. According to the results shown in table 4, the amount of pyruvic acid in fresh (sliced and crushed) in Trt 1(20 °C, 0 min storage) is low, and reached its highest value in in Trt 2 (4 °C, 10 min storage), and then begins to gradually decrease with the passage of time and the increase in temperature, as confirmed by Metrani *et al.*, (2018). Concerning the pyruvic acid content in dried garlic, based on the results of our study, all treatments had a low percentage of pyruvic acid content and this decrease might caused by the thermal process. Loss of pungency in garlic may be caused by the destruction of precursors and partial inactivation of the enzyme allinase under drying conditions (Ratti *et al.*, 2007).

Total Phenol Content

The mean of total phenol content (TPC) in fresh (sliced and crushed) and dried garlic slices with the effect of different storage conditions (time and temperature) is given in tables 6 and 7. The total phenolic concentration of all dried treatments was considerably less than that of fresh treatments (sliced and crushed). Heat treatment might create permanent chemical changes and a reduction in total phenolic compounds since they are heat-labile (Djendoubi *et al.*, 2012). The TPC is affected by a number of variables, including temperature, light, water, and nutrient availability (Bystrická *et al.*, 2013). As shown in table 6 and 7, the result matches the study done by Pedisić *et al.*, (2018), who found that TPC in crushed garlic at room temperature for different exposure times (0, 2, 5, and 8 min) decreased. Given that TPC acts as an antioxidant, its possible oxidation to

quinones can be used to explain why TPC levels decreased during storage (Jastrzebski *et al.*, 2007). During processing and storage, many polyphenols are unstable and highly susceptible to oxidation and degradation (Chandra *et al.*, 2021). When plant cells are destroyed, the polyphenol oxidation and degradation reactions of polyphenols will be released and catalyzed, releasing the polyphenol oxidase, polyphenol peroxidase, polyphenol glycosidase, and polyphenol esterase in fruits and vegetables (Montoya *et al.*, 2021). This elucidates the rationale behind the elevated levels of total phenol content observed in whole garlic, followed by sliced garlic, and followed by crushed garlic.

Antioxidant Activity

The mean of antioxidant activity in fresh (sliced and crushed) and dried garlic slices with the effect of different storage conditions (time and temperature) is presented in tables 8 and 9. There were significant variances in antioxidant activity between garlic forms. Various parameters, such as heat treatment and extracted solvent, can alter the antioxidant activity of garlic (Songsungkan and Chanthai, 2014). The antioxidant activity comes from total phenol and organosulfur compounds (allyl cysteine, alliin, allicin, and allyl disulfide) in garlic. Total phenolic and antioxidant activity have a positive relationship (Kyung, 2012), and the thiosulfinates, in particular allicin, are capable of scavenging the peroxy radical and serving as antioxidants (Okada *et al.*, 2005). Temperature caused chemical oxidation and released oxidative and hydrolytic enzymes that might degrade the antioxidant chemicals in 46 vegetables, and air exposure also accelerated oxidation processes and enzyme activity (Altemimi *et al.*, 2017).

Sensory Evaluation

As the hedonic scale results showed in table 10, there were some significant differences ($P < 0.05$) between all samples for overall appearance, overall aroma, overall pungency, and overall flavor. while there were no differences between samples for overall color compared to the control. The results of consumer testing as hedonic scale showed that the highest value of overall appearance was found in sample C3 'hummus with crushed garlic at 10 min 20 °C' (7.63), the highest value of overall aroma found in sample C3 'hummus with crushed garlic at 10 min 20 °C' (7.63), the highest value of overall flavor found in sample C3 'hummus with crushed garlic at 10 min 20 °C' (7.53), the highest value of overall pungency found in sample C3 'hummus with crushed garlic at 10 min 20 °C' (7.63). Table 11 shows the just-about-right scale according to flavor, aroma, and pungency. The highest value of flavor, aroma, and pungency are found in sample D4 'hummus with dried garlic at 20 min 4 °C', while the lowest value was found in 'hummus with crushed garlic at 10 min 4 °C' (1.13). These data also agreed with Al-Nabulsi *et al.*, (2022), who found that hummus without garlic was rated 6.9 in color, 6.9 in aroma, 6.9 in overall flavor, and 6.7 in Overall impression. In the same study, when adding 1% garlic to hummus, the sample was rated 6.9 in color, 5.9 in aroma, 5.9 in overall flavor, and 4.5 in the overall impression.

CONCLUSIONS

This study investigated different forms of garlic (freshly sliced, freshly crushed, and dried slices). The rate of allicin formation was significantly affected by temperature, and fresh mashed garlic had more content of allicin and pyruvic acid than fresh sliced garlic. On the other hand, fresh sliced garlic has a higher content of allicin and pyruvic acid than dried garlic slices. Optimum time and temperature in which a large amount of allicin is obtained after 10 minutes of mashing fresh garlic

273 at room temperature. Total phenol content, antioxidant activity, pyruvic acid, and allicin decrease
274 with increased time and temperature. For sensory attributes fresh mashed garlic after 10-minute
275 storage at 20 °C have the most acceptable pungency attributes.

276 **Data availability:**

277 Data is available at the following link: <https://data.mendeley.com/datasets/scgr52tw9h/1>.

278 **Acknowledgment**

279 The support provided by the Deanship of Research (128-2018) at Jordan University of Science
280 and Technology is appreciated. The authors extend their appreciation to the Researchers
281 Supporting Project number (RSP2023R502), King Saud University, Riyadh, Saudi Arabia for
282 funding this project. The support that was provided by the Deanship of Research at Jordan
283 University of Science and Technology is appreciated

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285 **REFERENCE**

- 286 Abe, K., Hori, Y., and Myoda, T. (2020). Characterization of key aroma compounds in aged garlic
287 extract. In: Food Chemistry, 312(2), 126081. <https://doi.org/10.1016/j.foodchem.2019.126081>
- 288 Al-Nabulsi, A. A., Hasan, F., Olaimat, A. N., Taha, S., Ayyash, M., Nazzal, D. S., Savvaidis, I.
289 N., Obaid, R. S., and Holley, R. (2022). Antimicrobial effects of chitosan and garlic against
290 salmonella spp., escherichia coli O157:H7, and listeria monocytogenes in hummus during storage
291 at various temperatures. In: Journal of Food Science, 87(2), 833–844.
292 <https://doi.org/10.1111/1750-3841.16025>

293 Altemimi, A., Lakhssassi, N., Baharlouei, A., Watson, D., and Lightfoot, D. (2017).
294 Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant
295 extracts. In: Plants, 6(4), 42. <https://doi.org/10.3390/plants6040042>

296 Alu'datt, M. H., Alli, I., Ereifej, K., Alhamad, M., Al-Tawaha, A. R., and Rababah, T. (2010).
297 Optimisation, characterisation and quantification of phenolic compounds in olive cake. In: Food
298 Chemistry, 123(1), 117-122

299 Amagase, H. (2006). Clarifying the real bioactive constituents of garlic. In: The Journal of
300 Nutrition, 136(3), 112. <https://doi.org/10.1093/jn/136.3.716s>

301 Ammarellou, A., Yousefi, A. R., Heydari, M., Uberti, D., and Mastinu, A. (2022). Biochemical
302 and botanical aspects of allium sativum L. sowing. In: BioTech, 11(2), 16.
303 <https://doi.org/10.3390/biotech11020016>

304 Anthon, G. E., and Barrett, D. M. (2003). Modified method for the determination of pyruvic acid
305 with dinitrophenylhydrazine in the assessment of Onion Pungency. In: Journal of the Science of
306 Food and Agriculture, 83(12), 1210–1213. <https://doi.org/10.1002/jsfa.1525>

307 Bat-Chen, W., Golan, T., Peri, I., Ludmer, Z., and Schwartz, B. (2010). Allicin purified from fresh
308 garlic cloves induces apoptosis in colon cancer cells via NRF2. In: Nutrition and Cancer, 62(7),
309 947–957. <https://doi.org/10.1080/01635581.2010.509837>

310 Bhandari, P. R. (2012). Garlic (allium sativum L.): A review of potential therapeutic applications.
311 In: International Journal of Green Pharmacy, 6(2), 118. <https://doi.org/10.4103/0973-8258.102826>

312 Botas, J., Fernandes, Â., Barros, L., Alves, M. J., Carvalho, A. M., and Ferreira, I. C. F. R. (2019).
 313 A comparative study of black and white allium sativum L.: Nutritional composition and bioactive
 314 properties. In: *Molecules*, 24(11), 2194. <https://doi.org/10.3390/molecules24112194>

315 Brand-Williams, W., Cuvelier, M. E. and Berset, C. (1995). Use of a free radical method to
 316 evaluate antioxidant activity. In: *LWT - Food Science and Technology*, 28(1), 25–30. doi:
 317 10.1016/S0023-6438(95)80008-5

318 Bystrická, J., Musilová, J., Vollmannová, A., Timoracká, M., and Kavalcová, P. (2013). Bioactive
 319 components of onion (*allium cepa*L.) — a review. In: *Acta Alimentaria*, 42(1), 11–22.
 320 <https://doi.org/10.1556/aalim.42.2013.1.2>

321 Chan, J. Y.-Y., Yuen, A. C.-Y., Chan, R. Y.-K., and Chan, S.-W. (2012). A review of the
 322 cardiovascular benefits and antioxidant properties of allicin. *Phytotherapy Research*, 27(5), 637–
 323 646. <https://doi.org/10.1002/ptr.4796>

324 Chandra, R. D., Prihastyanti, M. N. U., and Lukitasari, D. M. (2021). Effects of pH, high pressure
 325 processing, and ultraviolet light on carotenoids, chlorophylls, and anthocyanins of fresh fruit
 326 and vegetable juices. In: *eFood*, 2(3), 113–124

327 De Diego, M., Avello, M., Mennickent, S., Fernández, M., and Fernández, P. (2007). Validated
 328 liquid chromatographic method for quantitative determination of allicin in garlic powder and
 329 tablets. In: *Journal of separation science*, 30(16), 2703-2707

330 Djendoubi Mrad, N., Boudhrioua, N., Kechaou, N., Courtois, F., and Bonazzi, C. (2012). Influence
 331 of air drying temperature on kinetics, physicochemical properties, total phenolic content and

332 ascorbic acid of pears. In: Food and Bioproducts Processing, 90(3), 433–441.
333 <https://doi.org/10.1016/j.fbp.2011.11.009>

334 Fariás-Campomanes, A. M., Horita, C. N., Pollonio, M. A., and Meireles, M. A. A. (2014). Allicin-
335 rich extract obtained from garlic by pressurized liquid extraction: Quantitative determination of
336 allicin in garlic samples. In: Food and Public Health, 4(6), 272-278

337 Fujisawa, H., Suma, K., Origuchi, K., Kumagai, H., Seki, T., and Ariga, T. (2008). Biological and
338 chemical stability of garlic-derived allicin. In: Journal of Agricultural and Food Chemistry, 56(11),
339 4229–4235. <https://doi.org/10.1021/jf8000907>

340 Gam, D.-H., Park, J.-H., Kim, J.-H., Beak, D.-H., and Kim, J.-W. (2021). Effects of allium sativum
341 stem extract on growth and migration in melanoma cells through inhibition of VEGF, MMP-2,
342 and MMP-9 genes expression. In: Molecules, 27(1), 21.
343 <https://doi.org/10.3390/molecules27010021>

344 Henriquez, C., Cordova, A., Almonacid, S. and Saavedra, J. (2014). Kinetic modeling of phenolic
345 compound degradation during drum-drying of apple peel by-products. In: J Food Eng. 143, 146–
346 153

347 Ilić, D. P., Stojanović, S., Najman, S., Nikolić, V. D., Stanojević, L. P., Tačić, A., and Nikolić, L.
348 B. (2015). Biological evaluation of synthesized allicin and its transformation products obtained by
349 microwaves in methanol: Antioxidant activity and effect on cell growth. In: Biotechnology &
350 Biotechnological Equipment, 29(1), 189–194. <https://doi.org/10.1080/13102818.2014.994267>

351 Jastrzebski, Z., Leontowicz, H., Leontowicz, M., Namiesnik, J., Zachwieja, Z., Barton, H.,
352 Pawelzik, E., Arancibia-Avila, P., Toledo, F., and Gorinstein, S. (2007). The bioactivity of

353 processed garlic (*allium sativum* L.) as shown in vitro and in vivo studies on Rats. In: Food and
 354 Chemical Toxicology, 45(9), 1626–1633. <https://doi.org/10.1016/j.fct.2007.02.028>

355 Khar, A., Banerjee, K., Jadhav, M. R., and Lawande, K. E. (2011). Evaluation of garlic ecotypes
 356 for allicin and other allyl thiosulphinates. In: Food Chemistry, 128(4), 988–996.
 357 <https://doi.org/10.1016/j.foodchem.2011.04.004>

358 Kovarovič, J., Bystrická, J., Vollmannová, A., Tóth, T., and Brindza, J. (2019). Biologically
 359 valuable substances in garlic (*allium sativum* L.) – a review. In: Journal of Central European
 360 Agriculture, 20(1), 292–304. <https://doi.org/10.5513/jcea01/20.1.2304>

361 Kyung, K. H. (2012). Antimicrobial properties of allium species. In: Current Opinion in
 362 Biotechnology, 23(2), 142–147. <https://doi.org/10.1016/j.copbio.2011.08.004>

363 Lawless, H. T., and Heymann, H. (2010). In: Sensory evaluation of food: principles and practices
 364 (Vol. 2). New York: Springer

365 Liu P, Weng R, Sheng X, Wang X, Zhang W, Qian Y, and Qiu J (2020) Profiling of organosulfur
 366 compounds and amino acids in garlic from different regions of China. In: Food Chem, 305,
 367 125499. <https://doi.org/10.1016/j.foodchem.2019.125499>

368 Marchese, A., Barbieri, R., Sanches-Silva, A., Daglia, M., Nabavi, S. F., Jafari, N. J., Izadi, M.,
 369 Ajami, M., and Nabavi, S. M. (2016). Antifungal and antibacterial activities of allicin: A Review.
 370 In: Trends in Food Science & Technology, 52, 49–56. <https://doi.org/10.1016/j.tifs.2016.03.010>

371 Martins, N., Petropoulos, S., and Ferreira, I. C. F. R. (2016). Chemical composition and bioactive
 372 compounds of garlic (*allium sativum* L.) as affected by pre- and post-harvest conditions: A
 373 Review. In: Food Chemistry, 211, 41–50. <https://doi.org/10.1016/j.foodchem.2016.05.029>

374 Metrani, R., Jayaprakasha, G. K., and Patil, B. S. (2018). Optimized method for the quantification
375 of pyruvic acid in onions by microplate reader and confirmation by High Resolution Mass Spectra.
376 In: Food Chemistry, 242, 451–458. <https://doi.org/10.1016/j.foodchem.2017.08.099>

377 Montoya, C. C., Valencia, W. G., Sierra, J. A., and Penagos, L. (2021). Enhanced pink-red hues
378 in processed powders from unfermented cacao beans. In: LWT, 138, 110671.
379 <https://doi.org/10.1016/j.lwt.2020.110671>

380 Netzel, M. E. (2020). Garlic: Much more than a common spice. In: Foods, 9(11), 1544.
381 <https://doi.org/10.3390/foods9111544>

382 Nguyen, B. T., Hong, H. T., O'Hare, T. J., Wehr, J. B., Menzies, N. W., and Harper, S. M. (2021).
383 A rapid and simplified methodology for the extraction and quantification of allicin in garlic. In:
384 Journal of Food Composition and Analysis, 104, 104114.
385 <https://doi.org/10.1016/j.jfca.2021.104114>

386 Okada, Y., Tanaka, K., Fujita, I., Sato, E., and Okajima, H. (2005). Antiodidant activity of
387 thiosulfinates derived from garlic. In: Redox Report, 10(2), 96–102.
388 <https://doi.org/10.1179/135100005x38851>

389 Olaimat, A. N., Al-Holy, M. A., Abu-Ghoush, M. H., Osaili, T. M., Al-Nabulsi, A. A., and Rasco,
390 B. A. (2017). Inhibition of *Shigella sonnei* and *Shigella flexneri* in hummus using citric acid and
391 garlic extract. In: Journal of food science, 82(8), 1908-1915

392 Pedisić, S., Zorić, Z., Miljanović, A., Šimić, D., Repajić, M., and Dragović-Uzelac, V. (2018).
393 Retention of bioactive compounds during domestic processing of Croatian domestic garlic (*allium*
394 *sativum* L.). In: Food Technology and Biotechnology, 56(4).
395 <https://doi.org/10.17113/ftb.56.04.18.5709>

396 Prati, P., Henrique, C. M., Souza, A. S., Silva, V. S., and Pacheco, M. T. (2014). Evaluation of
397 allicin stability in processed garlic of different cultivars. In: Food Science and Technology
398 (Campinas), 34(3), 623–628. <https://doi.org/10.1590/1678-457x.6397>

399 Rahman, M. S. (2007). Allicin and other functional active components in garlic: Health benefits
400 and bioavailability. In: International Journal of Food Properties, 10(2), 245–268.
401 <https://doi.org/10.1080/10942910601113327>

402 Rahman, M. S., Al-Shamsi, Q. H., Bengtsson, G. B., Sablani, S. S., and Al-Alawi, A. (2009).
403 Drying kinetics and allicin potential in garlic slices during different methods of drying. In: Drying
404 Technology, 27(3), 467–477. <https://doi.org/10.1080/07373930802683781>

405 Ratti, C., Araya-Farias, M., Mendez-Lagunas, L., and Makhlof, J. (2007). Drying of garlic (*allium*
406 *sativum*) and its effect on allicin retention. In: Drying Technology, 25(2), 349– 356.
407 <https://doi.org/10.1080/07373930601120100>

408 Rybak, M. E., Calvey, E. M., and Harnly, J. M. (2004). Quantitative determination of allicin in
409 garlic: supercritical fluid extraction and standard addition of Alliin. In: Journal of Agricultural and
410 Food Chemistry, 52(4), 682–687. <https://doi.org/10.1021/jf034853x>

411 Shang, A., Cao, S.-Y., Xu, X.-Y., Gan, R.-Y., Tang, G.-Y., Corke, H., Mavumengwana, V., and
412 Li, H.-B. (2019). Bioactive compounds and biological functions of garlic (*allium sativum* L.). In:
413 Foods, 8(7), 246. <https://doi.org/10.3390/foods8070246>

414 Songsungkan, J., and Chanthai, S. (2014). Determination of synergic antioxidant activity of the
415 methanol/ethanol extract of allicin in the presence of total phenolics obtained from the garlic
416 capsule compared with fresh and baked garlic clove. In: International Food Research Journal,
417 21(6), 2377

418 Thuwapanichayanan, R., Prachayawarakorn, S., and Soponronnarit, S. (2014). Heat and moisture
 419 transport behaviour and quality of chopped garlic undergoing different drying methods. In: Journal
 420 of Food Engineering, 136, 34–41. <https://doi.org/10.1016/j.jfoodeng.2014.03.017>
 421 Veríssimo, T., Almeida, I., Cidade, H., Pinto, M., Azevedo, S., Oliveira, B., and Cunha, L.M.
 422 (2010). Evaluation of antioxidant activity of minimally processed garlic cloves. In: XXVIII
 423 International Horticultural Congress - IHC2010, 77-178
 424 Yilmaz, E., and Aydeniz, B. (2012). Sensory evaluation and consumer perception of some
 425 commercial green table olives. In: British Food Journal, 114(8), 1085-1094
 426 Yusuf, A., Fagbuaro, S. S., and Fajemilehin, S. O. (2018). Chemical Composition, phytochemical
 427 and mineral profile of garlic (*allium sativum*). In: Journal of Bioscience and Biotechnology
 428 Discovery, 3(5), 105–109. <https://doi.org/10.31248/jbbd2018.073>
 429 Zalepugin, D. Yu., Tilkunova, N. A., & Chernyshova, I. V. (2015). Stability of thiosulfinates from
 430 garlic (*Allium sativum* L.) supercritical extracts in polar and nonpolar solvents. *Russian Journal*
 431 *of Physical Chemistry B*, 9(7), 1032–1042. <https://doi.org/10.1134/s1990793115070143>
 432 Zhou, L., Guo, X., Bi, J., Yi, J., Chen, Q., Wu, X., and Zhou, M. (2016). Drying of garlic slices
 433 (*allium sativum*L.) and its effect on thiosulfinates, total phenolic compounds and antioxidant
 434 activity during infrared drying. In: Journal of Food Processing and Preservation, 41(1).
 435 <https://doi.org/10.1111/jfpp.12734>
 436 Zhu, X.-Y., and Zeng, Y.-R. (2020). Garlic extract in prosthesis-related infections: A literature
 437 review. In: Journal of International Medical Research, 48(4), 030006052091377.
 438 <https://doi.org/10.1177/0300060520913778>